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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
09/530,907 06/30/2000		RUDI WILFRIED JAN PAUWELS	07619.0006	4853
45511	7590 12/21/2005		EXAMINER	
	K WASHBURN LLP	SHIBUY		, MARK LANCE
ONE LIBERTY PLACE 46TH FLOOR			ART UNIT	PAPER NUMBER
	HIA, PA 19103		1639	

DATE MAILED: 12/21/2005

Please find below and/or attached an Office communication concerning this application or proceeding.

,		Application No.	Applicant(s)			
		09/530,907	PAUWELS ET AL.			
	Office Action Summary	Examiner	Art Unit			
		Mark L. Shibuya	1639			
Period fo	The MAILING DATE of this communication app or Reply	pears on the cover sheet with the c	orrespondence address			
A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION. - Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication. - If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication. - Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).						
Status						
1)[🔀	1)⊠ Responsive to communication(s) filed on <u>14 September 2005</u> .					
′=		action is non-final.				
,	3) Since this application is in condition for allowance except for formal matters, prosecution as to the merits is					
,—	closed in accordance with the practice under <i>Ex parte Quayle</i> , 1935 C.D. 11, 453 O.G. 213.					
Disposit	ion of Claims		ĺ			
•		s/are pending in the application				
- ال	4) Claim(s) 1,5,10,17,18,24,26,29,30 and 32-35 is/are pending in the application. 4a) Of the above claim(s) is/are withdrawn from consideration.					
5)□	5) Claim(s) is/are allowed.					
, —	6)⊠ Claim(s) <u>1.5,10,17,18,24,26,29,30 and 32-35</u> is/are rejected.					
· · · · · · · · · · · · · · · · · · ·	7) Claim(s) is/are objected to.					
•	8) Claim(s) are subject to restriction and/or election requirement.					
Applicat	ion Papers					
		ne.				
9) The specification is objected to by the Examiner.						
10) The drawing(s) filed on is/are: a) accepted or b) objected to by the Examiner. Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).						
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.03(a).						
11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.						
Priority under 35 U.S.C. § 119						
	12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).					
a) ☐ All b) ☐ Some * c) ☐ None of: 1.☐ Certified copies of the priority documents have been received.						
	Certified copies of the priority documents have been received in Application No					
	3. Copies of the certified copies of the priority documents have been received in this National Stage					
application from the International Bureau (PCT Rule 17.2(a)).						
* See the attached detailed Office action for a list of the certified copies not received.						
Attachmen	ıt(s)					
1) 🔯 Notic	e of References Cited (PTO-892)	4) Interview Summary				
	ce of Draftsperson's Patent Drawing Review (PTO-948)	Paper No(s)/Mail Da 5) Notice of Informal P	ate ratent Application (PTO-152)			
	mation Disclosure Statement(s) (PTO-1449 or PTO/SB/08) er No(s)/Mail Date	6) Other:				

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DETAILED ACTION

1. Claims 1, 5, 10, 17, 18, 24, 26, 29, 30, and 32-35 are pending and examined.

2. Applicant's description of the status of the claims, (Reply at p. 15, para 2) does not comport with applicant's submission of the amended claims, entered 9/14/2005. Therefore, the examiner has taken the amended claims, entered 9/14/2005, as replacing all prior versions and any other listings.

Withdrawn Rejections

- 3. The rejection of Claims 2-4, 6, 7, 17-19, 26-28 and 31, under 35 U.S.C. 112, second paragraph, is withdrawn over applicant's arguments and amendments to the claims.
- 4. The rejection of Claims 1, 5-7, 9, 10, 17, 24, 26, 29, 30, and 32-36 under 35 U.S.C. 102(b) as being anticipated by Lerner et al. (US 5,601,992), is withdrawn in view of applicant's arguments and amendments to the claims.

Priority

5. The instant application is the national stage of PCT/IB98/01399, filed 9/8/1999.

New Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless -

- (e) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.
- 6. Claims 1, 5, 10, 17, 18, 24, 26, 29, 30, and 32-35 are rejected under 35 U.S.C. 102(e) as being anticipated by Bjornson et al., (US 6,284,113 B1, priority to 19 September 1997, of record). This rejection is necessitated by applicant's amendments to the claims, entered 9/14/2005.

The claims are drawn to methods for screening for analytes comprising the steps of a) disposing a plurality of analytes to be screened within individually identifiable containers such that the analytes remain isolated from each other, wherein the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array: b) dispensing the analytes through the open ends of the capillary tubes onto at least one solid support to maintain the transferred contents of each container separate from those of each other container, wherein said analytes are simultaneously applied onto the at least one solid support; (c) contacting said at least one analyte-carrying solid support with targets provided in a semi-solid or liquid medium, whereby said analytes are released from the at least one solid support to the targets, wherein each analyte when applied to the solid support

diffuses thereon so as to produce a concentration gradient; and (d) measuring analyte-target interactions, wherein said analyte-target interactions are measured using one or more of the following methods: microscopic, luminometric, densitometric, isotopic, and physical measurements; and variations thereof.

Bjornson et al., throughout the patent and abstract, teach methods wherein analytes or beads may be disposed within individually identifiable containers within microarray plates, and transferring the analytes or beads from the containers to microarray substrates in such a manner as to maintain the transferred contents of each container separate from those of each other container in the other microarray; wherein the microarrays comprise individually identifiable containers are in an array (e.g., col. 19, line 13; col. 20, line 13, Fig.s 6-8) of capillary tubes, including capillary tubes, and channels of capillary dimensions (col. 8, line 64-col. 9, line 4; col. 11, lines 6-10; col. 15, line 40-col. 16, line 67); wherein the microarray plates comprise individually identifiable cavity structures, reading on containers, and arrays of capillary channels, reading on capillary tubes, each of which is identifiable according to its position within the microfluidic network plate, and further comprising a array of microfluidic networks, and wherein transfer of the analytes to the at least one solid support occurs by dispensing thereof through apertures that are the open ends of the capillary channels (e.g., Fig. 4A, col. 16, lines 18-54); at col., e.g., col. 20, lines 46-53, teach microarray plates reading upon a solid support, wherein the microarray plates are information carriers which carry information in electronic and digitized form, (as in claim 18); wherein each analytebearing solid support is contacted in step b) with a target provided from a separate

compartment of a microarray plate that is a multi-compartmented apparatus; wherein said compartments are an arrangement of mini-wells in said apparatus; and at col. 9, lines 23-55, teach electroflow media, reading on solvents, wherein said media includes polysaccharides, agarose, natural and synthetic polymers such as methylcellulose, polyacrylamide, hydrogels.

Diffusion of analytes when applied to a solid support that is provided a gel, such as an agarose gel or a polymers hydrogel, whereupon a concentration gradient is formed, would be an inherent physical phenomenon, as evidenced by the publication of Gray et al., US 2005/0106380 A1. Gray et al. state:

Generally, diffusion may be either active and/or passive. In passive diffusion the molecule simply passes through a porous polymer opening in response to a concentration gradient and does not interact with the polymer. The passive diffusion rate is a function of polymer molecular size. For example, oxygen diffuses at a faster rate through LDPE than through HDPE. Passive diffusion is also a function of pore size and concentration gradient, and high exchange rates can occur with large pore size. Diffusion rate is also a function of the size of the diffusing molecule. For instance, for a given polymer, the diffusion rate for the following molecules is listed in the order of highest to lowest: oxygen, water, methanol and ethanol. Passive diffusion can be affected by factors such as polymer crosslinking and polymer elongation through stretching, vacuum packing or shrink wrapping. Generally passive permeability decreases with increasing degrees of crosslinking and elongation.

Gray et al., at p. 4, para [0031]. Thus, upon delivery of analytes to porous polymers, such as those taught by Bjornson et al., at col.s 9-10, passive diffusion of the analyte would inherently produce a concentration gradient, as in claim 1.

Bjornson et al., at col. 18, lines 24-61, especially lines 56-59; col. 30, line 56-col. 31, line 14, disclose the manipulation of bead and particles in the channels of their disclosed microfluidic arrays. Bjornson et al. at col. 29, line 66-col. 30, line 14, teach

using microfluidic processing for assays to determine specific binding pair members, as in determining an analyte, and include cell surface binding assays, assays for drug discovery and screening, and studies of receptors. Bjornson at, e.g., col. 28, lines 37-42, teach methods that employ detection means including, but not limited to spectrophotometric, chemiluminescent, electrochemical or radio chemical means. Bjornson at, e.g., col. 20, lines 46-65, teach the use of an electronic computer connected to electrodes, wherein the electrodes are interactive with an optical detection device such as ultraviolet or fluorescent spectrometer. Bjornson at, e.g., col. 24, lines 27-43, disclose dispensing liquid drops through capillary size dispensing tubes onto substrates surfaces in miniature arrays, where the printed arrays may consist of nucleic acids, peptides, immunoassay reagents, pharmaceutical test compounds and the like.

Maintained Claim Rejections - 35 USC § 103

The text of those sections of Title 35, U.S. Code not included in this action can be found in a prior Office action.

7. Claims 1-7, 9, 10, 17-19, 24, and 26-36 are rejected under 35 U.S.C. 103(a) as being unpatentable over Lerner et al., US 5,601,992, (of record) and Bjornson et al., (US 6,284,113 B1, priority to 19 September 1997, of record). This rejection is maintained for the reasons of record as set forth in the previous Office action. The rejection is copied below for the convenience of the reader.

The claims are drawn to methods for screening of analytes, comprising the steps of: a) simultaneously applying a plurality of analytes to be screened onto at least one solid support such that the analytes remain isolated from one another; b) contacting said at least one analyte-carrying solid

support with targets provided in a semi-solid or liquid medium, whereby said analytes are released from the at least one solid support to the targets; and c) measuring analyte-target interactions.

The claims are further drawn to methods according to Claim 1, wherein step (a) comprises (i) disposing the analytes within individually identifiable containers, and (ii) transferring the analytes from the containers to the at least one solid support in such a manner as to maintain the transferred contents of each container separate from those of each other container, (as in claim 2); wherein the individually identifiable containers are an array of capillary tubes, including capillary tubes, pens, including plotter pens, and print heads. (as in claim 3): wherein the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array, and wherein transfer of the analytes to the at least one solid support occurs by dispensing thereof through the open ends of the capillary tubes, (as in claim 4); wherein the solid support is an information carrier which carries information in electronic, magnetic or digitised form, (as in claim 18); wherein each analyte-bearing solid support is contacted in step b) with a target provided from a separate compartment of a multicompartmented apparatus, (as in claim 27); wherein said compartments are an arrangement of mini-wells in said apparatus, (as in claim 28); and wherein the analyte dissolves in a solvent, wherein said solvent includes gelatin, polysaccharides such as agar and agarose, natural and synthetic polymers such as methylcellulose, polyacrylamide, hydrogels, gels containing N-isopropylacrylamide, or thermo-sensitive polymers, such that each analyte following application to the solid support and drying, liquefies in response to said chemical or physical parameter, (as in claim 31). The claimed invention is interpreted in view of the rejection under 35 USC 112, Second Paragraph (discussed above in the instant Office action).

Lerner et al., discloses a method that reads on that of the instant claims. Specifically, the reference discloses detecting the interaction between an oligomeric molecule (reading on claimed analyte) and a target (see, e.g. Abstract). In the method of Lerner et al, a plurality of beads containing peptide analytes are applied to a substrate surface and allowed to diffuse therein (see, e.g. column 21, lines 33-66 – "[t]he oligomeric molecules diffuse through the substrate and interact with a target"). This reads on the claimed step b) of releasing the analytes from the solid supports. The reference also reads on step a) of having analytes on at least one solid support in an isolated fashion, see, for example, column 3, lines 5-22). Beads as solid supports are used for the peptide analytes and the interaction tests were run in culture dishes (see, e.g. Examples 1 & 3 of the reference), this reads on the supports recited in the instant claims. The culture dishes of the reference have gels thereon, see, for example, column 29, lines 62-66. This reads on a coated solid support as recited in the instant claims. The peptide analytes and their preparations (see, e.g. Example 1) read on the analytes recited in instant claims 29, 30 and 32. Various cellular targets are also described by the reference (see Example 2 and column 21, line 66 - column 22, line 67) reading on claim 33. In the reference, pigment dispersion is measured (see, e.g. column 25, line 55 - column 26, line 52); this reads on the limitations of instant claim 36.

Lerner et al. does not teach methods according to Claim 1, wherein step (a) comprises (i) disposing the analytes within individually identifiable containers, and (ii) transferring the analytes from the containers to the at least one solid support in such a manner as to maintain the transferred contents of each container separate from those of each other container. (as in claim 2); wherein the individually identifiable containers are an array of capillary tubes, including capillary tubes, pens, including plotter pens, and print heads, (as in claim 3); wherein the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array, and wherein transfer of the analytes to the at least one solid support occurs by dispensing thereof through the open ends of the capillary tubes, (as in claim 4); wherein the solid support is an information carrier which carries information in electronic, magnetic or digitised form, (as in claim 18); wherein each analyte-bearing solid support is contacted in step b) with a target provided from a separate compartment of a multicompartmented apparatus, (as in claim 27); wherein said compartments are an arrangement of mini-wells in said apparatus, (as in claim 28); and wherein the analyte dissolves in a solvent, wherein said solvent includes gelatin, polysaccharides such as agar and agarose, natural and synthetic polymers such as methylcellulose, polyacrylamide, hydrogels, gels containing N-isopropylacrylamide, or thermo-sensitive polymers, such that each analyte following application to the solid support and drying, liquefies in response to said chemical or physical parameter, (as in claim 31).

Bjornson et al., throughout the patent and abstract, teach methods wherein analytes or beads may be disposed within individually identifiable containers within microarray plates, and transferring the analytes or beads from the containers to microarray substrates in such a manner as to maintain the transferred contents of each container separate from those of each other container in the other microarray, (as in claim 2); wherein the microarrays comprise individually identifiable containers are in an array (e.g., col. 19, line 13; col. 20, line 13, Fig.s 6-8) of capillary tubes, including capillary tubes, and channels of capillary dimensions (col. 8, line 64-col. 9, line 4; col. 11, lines 6-10; col. 15, line 40-col. 16, line 67), (as in claim 3); wherein the microarray plates comprise individually identifiable cavity structures, reading on containers, and arrays of capillary channels, reading on capillary tubes, each of which is identifiable according to its position within the microfluidic network plate, and further comprising a array of microfluidic networks, and wherein transfer of the analytes to the at least one solid support occurs by dispensing thereof through apertures that are the open ends of the capillary channels (e.g., Fig. 4A, col. 16, lines 18-54), (as in claim 4); at col., e.g., col. 20, lines 46-53, teach microarray plates reading upon a solid support, wherein the microarray plates are information carriers which carry information in electronic and digitized form, (as in claim 18); wherein each analyte-bearing solid support is contacted in step b) with a target provided from a separate compartment of a microarray plate that is a multi-compartmented apparatus, (as in claim 27); wherein said compartments are an arrangement of mini-wells in said apparatus, (as in claim 28); and at col. 9, lines 23-55, teach electroflow media, reading on solvents, wherein said media includes polysaccharides, agarose, natural and synthetic polymers such as methylcellulose, polyacrylamide, hydrogels. Bjornson et al., at col. 18, lines 24-61, especially lines 56-59; col. 30, line 56-col. 31, line 14, disclose the manipulation of bead and particles in the channels of their disclosed microfluidic arrays. Bjornson et al. at col. 29, line 66-col. 30, line 14, teach using microfluidic processing for assay that determining specific binding pair members, as in determining an analyte, and including cell surface binding assays, assays for drug discovery and screening, and studies of receptors.

It would have been prima facie obvious at the time of the invention for one of ordinary skill in the art to have used methods comprising methods of screening of analytes, comprising applying a plurality of analytes onto solid supports, such that the analytes remain isolated from one another, and wherein the analytes are released after contact of the analyte-carrying solid supports; and wherein the analytes are disposed within individually identifiable containers, and transferring the analytes from the containers to the at least one solid support in such a manner as to maintain the transferred contents of each container separate from those of each other container, (as in claim 2); wherein the individually identifiable containers are an array of capillary tubes, (as in claim 3); wherein the individually identifiable containers are an array of capillary tubes each of which is identifiable according to its position within the array, and wherein transfer of the analytes to the at least one solid support occurs by dispensing thereof through the open ends of the capillary tubes, (as in claim 4); wherein the solid support is an information carrier which carries information in electronic or digitised form, (as in claim 18); wherein each analyte-bearing solid support is contacted in step b) with a target provided from a separate compartment of a multicompartmented apparatus, (as in claim 27); wherein said compartments are an arrangement of mini-wells in said apparatus, (as in claim 28); and wherein the analyte dissolves in a solvent, wherein said solvent includes polysaccharides, agarose, natural and synthetic polymers, methylcellulose, polyacrylamide, hydrogels, such that each analyte following application to the solid support and drying, liquefies in response to said chemical or physical parameter, (as in claim 31).

One of ordinary skill in the art would have been motivated to use methods of screening analytes, wherein the analytes are beads comprising oligomeric molecules, wherein the beads are applied to a substrate, as taught by Lerner (see, e.g., Lerner at col. 21, lines 33-43); and wherein the bead/analytes are manipulated in microarray plates that comprise compartments and capillary tubes, wherein the microarray plates are solid supports that are electronic, digital information carriers and further comprising media, as taught by Bjornson above, because Bjornson teaches manipulation of analytes or beads within microarray plates, and Bjornson teaches evaluating analyte binding to cell surfaces, for example, in order to identify specific binding pairs, and so to screen for potential drugs that target cell surface receptors.

One of ordinary skill in the art would have had a reasonable expectation of success in using bead analytes applied onto substrates, wherein those substrates are electronic microarrays comprising compartments, capillary tubes, and media, because, absent evidence to the contrary, beads or particles

that release compounds into solution were known in the medicinal arts and because flowing particles through such arrays, absent evidence to the contrary, were known in the microfluidic arts.

Applicant states in the Reply at p. 5, the language of cancelled claims 2-4, 9 and 36 was incorporated into amended claim 1. Because former claims 1-4, 9 and 36, were rejected as unpatentable over the teachings of the prior art references of Lerner and Bjornson, the instant rejection is maintained.

Applicant, in the Reply, reviews the previous Office action, and at p. 15, requests further clarification of the teachings of Bjornson et al., in regard to gathering data via an electronic, magnetic or digitized means. Applicant, in the Reply at pp. 16-17, argues that a *prima facie* case of obviousness has not been established, at least because there is no suggestion or motivation to modify the references; and applicant argues that the references when combined, do not teach or suggest all claim limitations of the currently pending, amended claims.

Response to Arguments

Applicant's arguments entered 9/14/2005 have been fully considered but they are not persuasive.

In regard to data collection, Bjornson at, e.g., col. 28, lines 37-42, teach assays and screening methods that employ detection means including, but not limited to spectrophotometric, chemiluminescent, electrochemical or radio chemical means.

Bjornson at, e.g., col. 20, lines 46-65, teach the use of an electronic computer connected to electrodes, wherein the electrodes are made interactive with an optical

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detection device, such as an ultraviolet or fluorescent spectrometer. Therefore, examiner respectfully submits that Bjornson teaches gathering data by electronic, magnetic or digitized means.

Bjornson et al., disclose motivation to combine the reference, as applicant notes in the Reply at p. 14, para 2, the previous Office action states:

One of ordinary skill in the art would have been motivated to use methods of screening analytes, wherein the analytes are beads comprising oligomeric molecules, wherein the beads are applied to a substrate, as taught by Lerner (see, e.g., Lerner at col. 21, lines 33-43); and wherein the bead/analytes are manipulated in microarray plates that comprise compartments and capillary tubes, wherein the microarray plates are solid supports that are electronic, digital information carriers and further comprising media, as taught by Bjornson above, because Bjornson teaches manipulation of analytes or beads within microarray plates, and Bjornson teaches evaluating analyte binding to cell surfaces, for example, in order to identify specific binding pairs, and so to screen for potential drugs that target cell surface receptors.

Office action, mailed 6/17/2005, at pp. 14-15, bridging paragraph. In particular, Bjornson at, e.g., col. 24, lines 27-43, disclose dispensing liquid drops through capillary size dispensing tubes onto substrates surfaces in miniature arrays, wherein the arrays may consist of nucleic acids, peptides, immunoassay reagents, pharmaceutical test compounds, and the like.

In response to applicant's argument that the examiner's conclusion of obviousness is based upon improper hindsight reasoning, it must be recognized that any judgment on obviousness is in a sense necessarily a reconstruction based upon hindsight reasoning. But so long as it takes into account only knowledge which was within the level of ordinary skill at the time the claimed invention was made, and does not include knowledge gleaned only from the applicant's disclosure, such a

reconstruction is proper. See *In re McLaughlin*, 443 F.2d 1392, 170 USPQ 209 (CCPA 1971).

As shown above, the examiner respectfully submits that motivation to combine the references may be found at least within the prior art reference of Bjornson et al. The examiner respectfully submits that applicant's arguments do not *refute* the aforementioned reasons for motivation to combine the references, as set forth in the previous Office action. Therefore, the examiner respectfully urges that a *prima facie* case of obviousness has, indeed, been established.

Conclusion

- 8. Claims 1, 5, 10, 17, 18, 24, 26, 29, 30, and 32-35 stand finally rejected.
- 9. Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of

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the advisory action. In no event, however, will the statutory period for reply expire later

than SIX MONTHS from the date of this final action.

10. Any inquiry concerning this communication or earlier communications from the

examiner should be directed to Mark L. Shibuya whose telephone number is (571) 272-

0806. The examiner can normally be reached on M-F, 8:30AM-5:00PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's

supervisor. Andrew Wang can be reached on (571) 272-0811. The fax phone number

for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the

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Business Center (EBC) at 866-217-9197 (toll-free).

Mark L. Shibuya Examiner

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